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Exploring the molecular basis for the metal-mediated assembly of alginate gels

Matthew B. Stewart[a,b,*], Stephen R. Gray[a,b], Todor Vasiljevic[a,c], John D. Orbell[a,b]

[a] Institute for Sustainability and Innovation (ISI), Victoria University, PO Box 14428, Melbourne, VIC 8001, Australia
[b] College of Engineering and Science, Victoria University, PO Box 14428, Melbourne, VIC 8001, Australia
[c] College of Health and Biomedicine, Victoria University, PO Box 14428, Melbourne, VIC 8001, Australia

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ABSTRACT

The binding of sodium and calcium ions to single and multiple poly-G decamer strands has been modelled by conducting a series of molecular dynamics simulations. Implications for metal mediated inter-strand interactions and gel assembly have been explored by systematically introducing up to three strands into each of these simulations. A particular emphasis has been placed on revealing intrinsic binding modes by an unbiased initial positioning of the metal ions. The results have revealed binding modes that provide a rationale for the observed gelling of alginate by calcium rather than sodium ions. A number of junction zones involving calcium ions have been identified that result in chain aggregation. This includes a distinctive perpendicular motif that appears to be ubiquitous in previously reported AFM images of open 3-D alginate networks. The coordination geometries of the metal ions have been characterised and the metal-mediated junctions between associated strands are described in detail.

1. Introduction

Alginate is the common term for a family of linear polyuronic acids isolated from brown algae (including several seaweed species) and some bacteria (Donati & Paoletti, 2009; Gorin & Spencer, 1966; Goven, Fyfe, & Jarman, 1981). Biologically, alginates are found in algal cell walls and intercellular mucilage, providing mechanical strength and flexibility – similar to the role played by cellulose and pectin in land-based plants (Donati & Paoletti, 2009). Alginates are also found in the protective cyst of *Azotobacter vilelandii* as well as in the biofilms produced by *Psuedomonas* and *Azotobacter* (Gorin & Spencer, 1966; Goven et al., 1981; Linker & Jones, 1966). Chemically, alginate chains consist of arrangements of β-D-mannuronic acid (M) and its C5 epimer, α-L-guluronic acid (G), bound *via* (1 → 4) glycosidic linkages, Fig. S1. Alginates are ionic at neutral pH, with a pKa around 3.8.

Isolated alginates (generally available as the Na^+ salt) are widely utilised throughout a number of industries, including as a common food additive and in a wide range of medical applications, from toothpastes to advanced wound dressings (BeMiller, 2009; Davis, Volesky, & Mucci, 2003; Donati & Paoletti, 2009; Langer & Tirrell, 2004). Such uses are generally related to their gelling and water-retaining properties. This sought-after gelling activity occurs when alginate chains aggregate in the presence of divalent cations (most commonly Ca^{2+}) that facilitate interchain interactions. Networks formed by such interchain aggregations may be visualised using techniques such as atomic force microscopy (Decho, 1999). We have relied upon the work of this author, in particular, in order to benchmark our MD results to experimental results. The areas where interchain interactions occur within these networks are commonly referred to as 'junction zones'. It has been shown previously that such junction zones may occur in G-rich areas of alginates and may display a preference for a free Ca^{2+}:G ratio of 1:4 (Grant, Morris, Rees, Smith, & Thom, 1973). An early attempt to define the specific nature of this interaction was the 'egg-box' model proposed by Grant and co-workers in 1973. This model proposed that Ca^{2+} binding occurs between two parallel alginate chains, which assume a 2_1 helical conformation. This model places the Ca^{2+} ions in pockets, or depressions, formed naturally by the puckered structure imparted by the G–G α-glycosidic bond, Fig. S1. Questions have been raised in the literature as to whether this structure is viable experimentally (Braccini & Pérez, 2001). Several other subsequent models have also been suggested, including the 'shifted egg-box' model (Braccini & Pérez, 2001) and a modified shifted egg-box model, whereby only carboxylate oxygen atoms interact with the Ca^{2+} ions (Plazinski, 2011). These previously published models of

* Corresponding author at: Institute for Sustainability and Innovation (ISI), Victoria University, PO Box 14428, Melbourne, VIC 8001, Australia. Tel.: +61 3 9919 7641; fax: +61 9919 8082.
E-mail addresses: Matthew.Stewart@vu.edu.au (M.B. Stewart), Stephen.Gray@vu.edu.au (S.R. Gray), Todor.Vasiljevic@vu.edu.au (T. Vasiljevic), John.Orbell@vu.edu.au (J.D. Orbell).

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Ca^{2+}–poly-G interactions have assumed that all Ca^{2+} interactions with G-rich sections involve parallel aggregation of the alginate chains and assume that there is only one mode of Ca^{2+} interaction. As noted previously in the literature (Plazinski, 2011), the precise environment of the Ca^{2+} ion in aggregated alginates is difficult to obtain using conventional analytical experimental techniques.

In this work, we have employed molecular dynamics methods to probe the binding of sodium and calcium ions to poly-G decamers in an aqueous environment – with a view to discerning preferred binding modes and their implications for interstrand interactions. Whilst the literature contains a number of previous theoretical studies which have attempted to provide insights into the specific binding site(s) of Ca^{2+} with alginates, this earlier work has been limited in scope and has tended to focus on confirming the widely accepted, if not entrenched, egg-box model (Braccini & Pérez, 2001; Li, Fang, Vreeker, & Appelqvist, 2007; Plazinski, 2011). The simulations carried out in the present work are designed to systematically characterise and compare the binding of charge-neutralising levels of Na^+ and Ca^{2+} ions to the poly-G sequence (represented here by one, two and three decamer chains) in an aqueous environment, and to examine the nature of possible metal-mediated interactions leading to aggregation between alginate strands. A particular goal has been to reveal *intrinsic* interactions, either of an intermediate or an essentially consolidated nature. In this regard, these simulations differ from previously published investigations (Braccini & Pérez, 2001; Braccini, Grasso, & Pérez, 1999; DeRamos, Irwin, Nauss, & Stout, 1997; Plazinski, 2011) in that the initial placement of the metal ions is random in relation to the alginate chains, rather than at specific distances between two parallel or anti-parallel chains of the poly-G. This is to ensure that any structures formed are not a function of the initial starting geometry. Thus the aim is not to replicate the classical 'egg-box' model, which is usually assumed to result in the metal-mediated parallel association of strands, but rather to delineate possible variants and alternatives that might also promote non-parallel arrangements or motifs that could form the basis for open grid-like configurations. Such arrangements could represent precursor structures for the formation of 3-D gel networks and could be suggestive of assembly mechanisms.

2. Computational method

Simulations were carried out using a modified CHARMM all22 force field (MacKerell et al., 1998) and the NAMD 2.7 package (Phillips et al., 2005). Topology and parameter files for carbohydrates were edited to allow for the addition of the carboxylic acid groups. Atomic point charges were recalculated from density functional equilibrium geometry calculations, at the B3LYP/6-31G(d) level of theory, on the G monomers and reapplied to the topology. Oligomers of the poly-G sequence, each 10 residues long and fully deprotonated, were constructed using the Visual Molecular Dynamics (VMD) program (Humphrey, Dalke, & Schulten, 1996) and optimised using MD in a TIP3P water box which measured 20 Å × 20 Å × 20 Å. These optimisations involved 2000 conjugate gradient minimisation steps, followed by 2 ps of molecular dynamics simulation time under NVT conditions in a periodic box. The temperature and pressure was maintained using Langevin dynamics as implemented in NAMD. It must be noted that these initial optimisation simulations were not charge neutralised, as the effect of metal ions (including Na^+, which is the standard counter ion for alginates) was to be investigated. These optimised structures were used as starting geometries for the metal ion binding simulations.

The production simulations involved either one, two or (in the case of the Ca^{2+} ion environments only) three oligomers. Where two oligomer chains were present, the identical sequences were placed parallel, approximately 6 Å apart; where three chains were used, these were placed in a triangular array. TIP3P water was then added, again with 20 Å padding in all directions, as was the case for the optimisation simulations above. Ca^{2+} ions were added randomly to the solvated system to neutralise the anionic charges of the oligomers. These ions were added *via* the Autoionize plugin in VMD with a minimum distance of 5 Å between the added ions and the solute or other ions. The ion placement was randomised so as to not bias the simulation towards a particular outcome. Each system was again subjected to 2000 conjugate gradient minimisation steps, followed by a simulated annealing procedure. This has been introduced in order to overcome the inherent energy barriers in these simulations whilst keeping the simulation time as short as possible. Achieving this by traditional optimisation methods requires the initial state of the system, in terms of the positioning of the Ca^{2+} ions, to reflect the expected outcome of the simulation – an approach that has been adopted in all previous MD studies on metal/alginate systems, *vide supra.* Since a major objective of this work is to avoid such bias, this simulated annealing protocol has been successfully implemented in order to readily identify more intrinsic outcomes. Notably, this approach has been validated in this study by the observed consolidation of well-behaved metal coordination geometries that are in good agreement with experimental data (Dudev & Lim, 2004; Einspahr & Bugg, 1981; Katz, Glusker, Beebe, & Bock, 1996). A major advantage of this method is that highly informative structural outcomes are obtained in a relatively short simulation time of less than 1 ns. Specifically, the annealing procedure involved an initial increase in temperature to 650 K, with cooling of the system to 300 K in 50 K increments at 100 ps intervals, giving a final runtime of 0.8 ns. An NVT-ensemble and periodic boundary conditions were utilised for these annealing simulations with the temperature controlled *via* rescaling of all velocities every 2 ps. The equations of motion were integrated at 1 fs timesteps. Short-range nonbonded interactions were calculated at a 1 ps timestep with a Van der Waals switching distance of 10 Å and overall nonbonded cutoff at 12 Å with long-range interactions integrated every 2 ps using the r-RESPA integrator. The Particle mesh Ewald (PME) method was used to describe the full-system periodic electrostatics. The short-range pairlist was updated every 10 ps for atoms within 14 Å. The simulation trajectory was written every 500 fs in order to provide good quality data with respect to interactions. All single chain simulation trajectories were analysed *via* calculation of the radial distribution functions for specific oxygen–metal ion interactions, utilising the $g(r)$ GUI plugin of VMD (Humphrey et al., 1996).

3. Convention

The numbering scheme for the G monomer with respect to the oxygen atoms, and the convention for referring to these atoms at different locations in the chain, is given in Fig. 1.

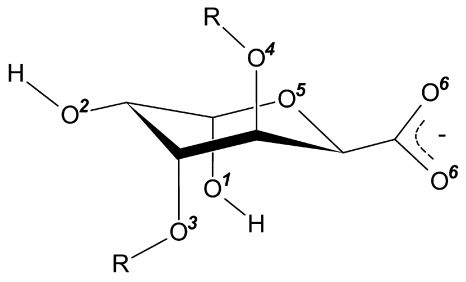

Fig. 1. The G alginate monomer, α-L-guluronate, showing the numbering scheme with respect to the oxygen atoms. Note: atoms referred to in the text that are in an adjacent monomer are marked with a dash; a double dash refers to atoms located two monomers away; an asterisk indicates that an atom is on a second chain; O6 refers to the oxygens of a delocalized carboxylate anion.

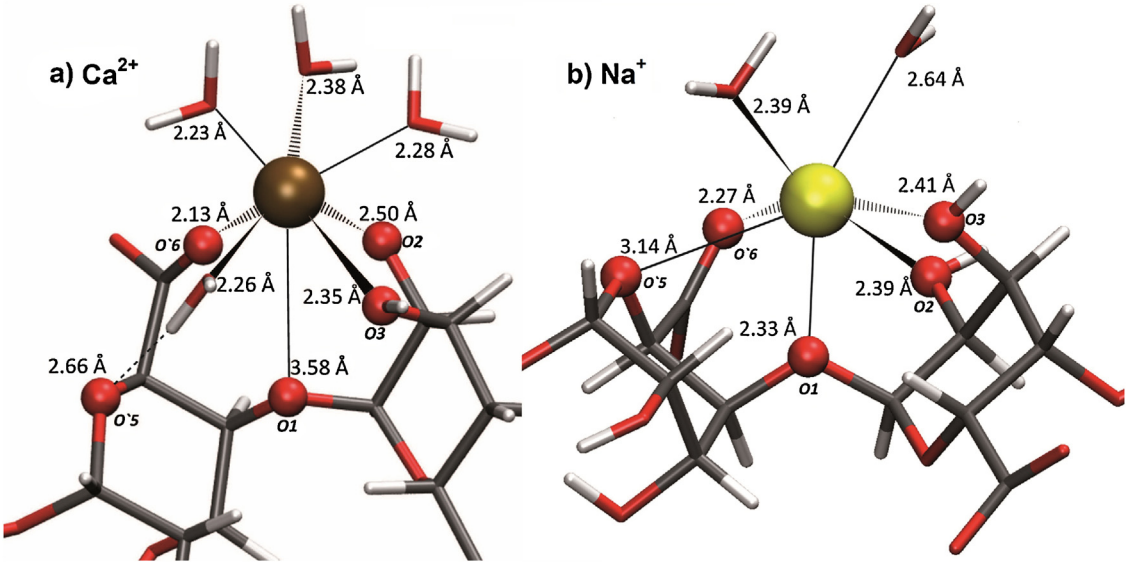

Fig. 2. (a) The egg-box binding site for Ca^{2+} (brown sphere); note the water bridge to the O'5 atom. (b) The egg-box binding site for Na^{+} (yellow sphere). In (a) and (b) oxygen atoms on the poly-G chain are shown as red spheres; coordinated water molecules are shown as stick formulae. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Simulations involving a single alginate decamer are denoted by 1 x poly-G, those involving two such chains are denoted by 2 x poly-G and those involving three such chains are denoted by 3 x poly-G.

4. Results and discussion

The metal ion oligomer interactions observed in this study are generally formed rapidly – with Ca^{2+} interactions consolidating sooner within the simulations than Na^{+}. Notably, chains were observed to aggregate only in the presence of Ca^{2+} ions. This is consistent with experimental observations regarding the solubility of alginates in the presence of monovalent cations and hydrogel formation in the presence of divalent or multivalent cations (Donati & Paoletti, 2009). It is acknowledged that this study relates to decamers only and may not directly reveal the influence of chain length on the longer-term aggregation dynamics (Kohn, 1975). However, this work has been designed to specifically investigate the *initial* stages of aggregation, the motifs of which may then form the basis for subsequent cooperative aggregation. The detailed interactions of Ca^{2+} and Na^{+} with the poly-G decamer are described as follows:

4.1. 1 x Poly-G

When a single poly-G chain, in the presence of water and Ca^{2+} ions is simulated, two distinct binding interactions are identified. The most prevalent interaction, which involves four of the five Ca^{2+} ions available in this simulation, involves direct binding to carboxylate groups. These Ca^{2+} ions are each coordinated to one carboxylate oxygen and between five and six water molecules. A consistent feature of these interactions is the hydrogen bonding that is observed to occur between one of the coordinated water molecules and the O3 atom of the neighbouring guluronate residue, Fig. S2. Such 'water bridging' would be expected to stabilise such interactions. The other identified interaction is similar, although not exactly identical to, the classical egg-box model, Fig. 2. This interaction involves four coordinating oxygen atoms from the poly-G, reflecting the egg-box model, and four coordinated water molecules. Whereas the classical egg-box model postulates five coordinating atoms per alginate chain, specifically O'6, O'5, O2, O3 and O1, the interaction identified in this study forms through direct coordination with the O'6, O2, O3 and O1 atoms only – with one of the coordinated water molecules forming a water bridge to the O'5 atom. For this binding mode, Fig. 3a plots the distance of the coordinating oxygen atoms from the Ca^{2+} ion as a function of the simulation time. It can be seen from Fig. 3a that the interaction distances appear to be relatively stable from approximately 0.4 ns onwards, which corresponds to an annealing temperature of simulation of 450 K. Only three oxygen atoms of the guluronate (O'6, O2 and O3) can be considered to be tightly bound to the Ca^{2+} – with average bound distances of 2.16 ± 0.7 Å, 2.45 ± 0.16 Å, and 2.42 ± 0.17 Å, respectively.

The O1 atom exhibits only a weak interaction with the Ca^{2+} – at an average distance of 3.23 ± 0.5 Å and the O'5 atom is not considered to be close enough to the Ca^{2+} ion to exert a direct influence – with an average distance of 4.13 ± 0.6 Å. This coordination geometry does not strictly agree with the classical egg-box model, which proposes that all five oxygen atoms described here interact directly with the Ca^{2+} ion (Grant et al., 1973). Rather, it is in closer agreement with the revisited egg-box model of Braccini and Pérez, where interactions with O2 and O3 are also identified, and which does not attribute a role to the O'5 oxygen atom (Braccini & Pérez, 2001). Thus the interaction observed here can be regarded as a hybrid of these two models. Whilst only four oxygen atoms are directly interacting with the Ca^{2+} ions, the O'5 atom interacts with the Ca^{2+} ion *via* a coordinated water molecule (water bridging), Fig. 2a.

Fig. 4 shows the corresponding radial pair distribution function, $g(r)$, for Ca^{2+} *versus* the five coordinating alginate oxygen atoms.

This plot clearly shows that Ca^{2+} displays a high preference (*i.e.* interaction probability – $g(r)$-O'6 is plotted separately on the right hand vertical axis) for binding to the negatively charged carboxylate moiety, O'6; at about 2.13 Å. O2 and O3 also bind directly to Ca^{2+} at about 2.5 Å, which is consistent with the data presented in Fig. 3. The O'5 atom shows no direct interaction with the Ca^{2+} in the $g(r)$ plot (*i.e.* less than approximately 3 Å). However, it does show some interaction with Ca^{2+} occurring at distances between 4.3 and 5 Å, *via* a water-bridging interaction.

When this same single chain system is simulated in the presence of Na^{+} ions, there is a markedly different interaction profile compared to Ca^{2+}. The dominant interaction in this simulation is the Na^{+} equivalent to the classical egg-box structure as previously

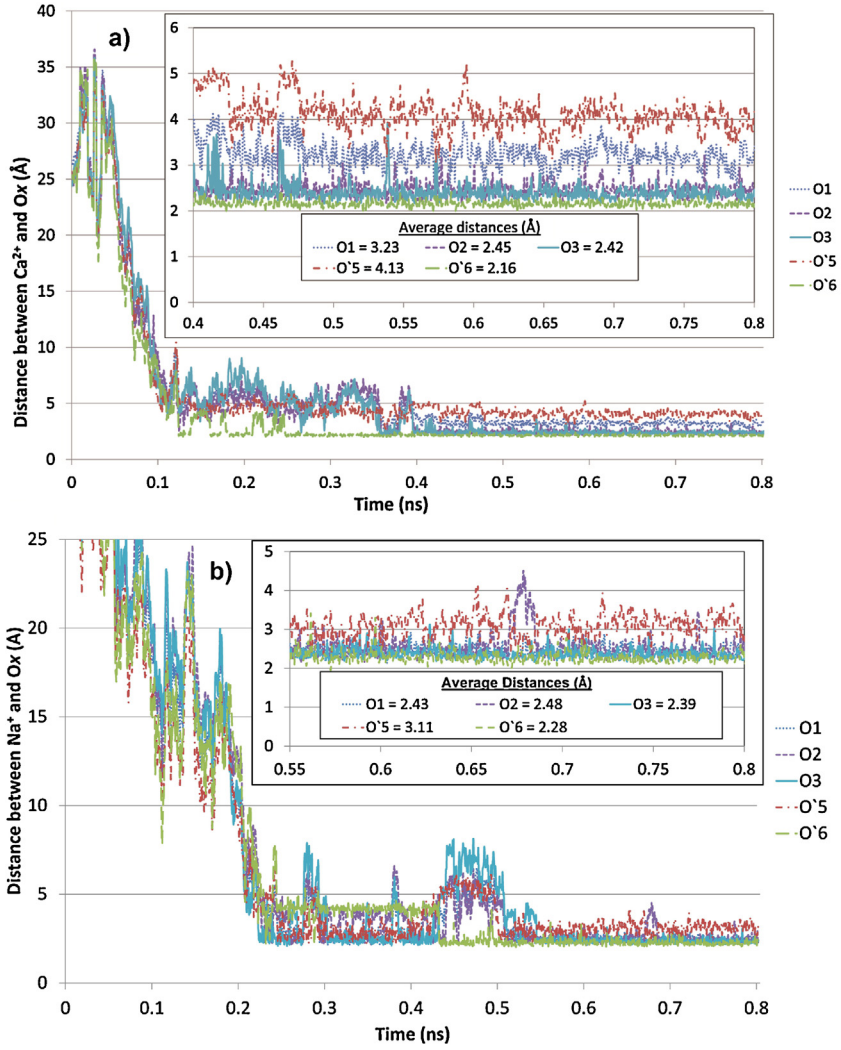

Fig. 3. (a) The distances between the Ca^{2+} ion and the bonding oxygen atoms in the variant of the egg-box structure as identified in the single poly-G chain simulation. Note the apparent stability in these distances from approximately 0.4 ns – suggesting a consolidation of the interaction. (b) Distances from the coordinating Na^{+} ion with the five interacting oxygen atoms of the single poly-G chain at the binding site between residues 3 and 4 of the decamer. The inset plots are magnifications of the last 0.4 and 0.25 ns respectively.

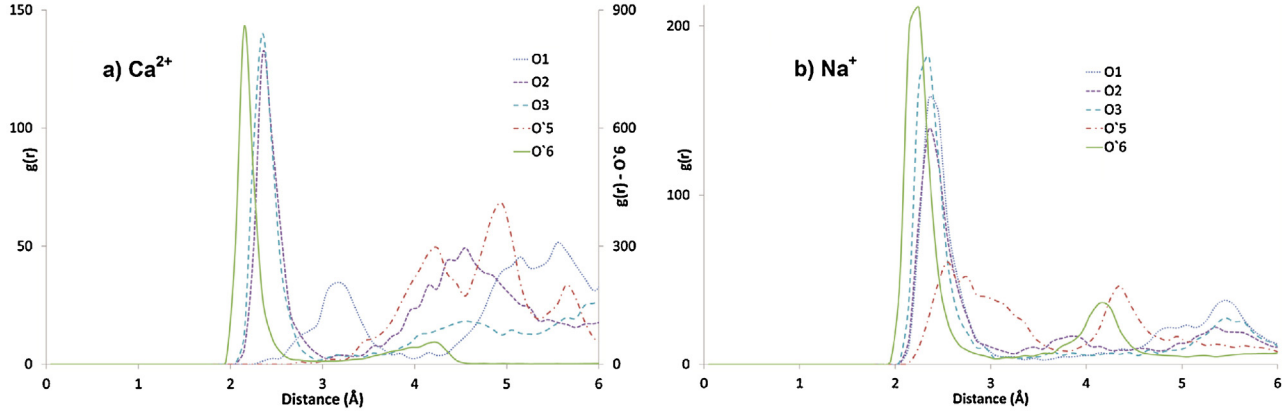

Fig. 4. Comparative radial distribution functions, $g(r)$, for (a) Ca^{2+} and (b) Na^{+} against each of the oxygen atoms of interest in the poly-G sequence. In (a), the $g(r)$ function values for the O‘6 atom are plotted on the secondary axis.

postulated for Ca^{2+}, Fig. 2b. To the best of our knowledge, this has not been previously characterised in the literature.

In this simulation, two such binding sites are found that have Na^{+} directly coordinated to the O‘6, O‘5, O2, O3 and O1 atoms (as required by the classical Ca^{2+}-based egg-box model) whilst the third binding site does not fully coordinate the O‘5 atom, Fig. 2b. However, as this occurs in a terminal residue, there is expected to be additional conformational flexibility at this site (*i.e.* an ‘end

Table 1
Comparison of the average angles and distances involved in the calcium and sodium egg-box structures.

	M = Sodium	M = Calcium	Δ (Ca–Na)
Angle			
O2-M-O'6	151.00°	124.12°	−26.88°
O1-M-O'6	82.04°	61.82°	−20.22°
O1-M-O2	70.11°	57.93°	−12.18°
O1-M-O3	76.21°	63.43°	−12.78°
Distance			
M-O1	2.43 Å	3.23 Å	0.8 Å
M-O2	2.48 Å	2.45 Å	−0.03 Å
M-O3	2.39 Å	2.42 Å	−0.03 Å
M-O'5	3.11 Å	4.13 Å	1.02 Å
M-O'6	2.28 Å	2.16 Å	−0.12 Å

effect'). There are also two identified occurrences of water-bridged Na^+-carboxylate interactions analogous to those identified for Ca^{2+}, Fig. S2; with the other five Na^+ ions present in the simulation not interacting with the chain. The relative proportions of these interactions varied over the simulation time, as the interactions between the Na^+ and poly-G chain were not as stable as for the Ca^{2+} simulation, with a higher degree of reversibility of the binding modes. This is evident in Fig. 3b that plots the Na^+-oxygen distances for one of the Na^+ binding sites identified in this simulation. When compared to Fig. 3b, the radial pair distribution function, $g(r)$, for Na^+, Fig. 4b, highlights the preference of this ion for the full (classical) egg-box binding with the atoms of the coordination sphere, including the carboxylate, showing roughly equivalent probabilities of interaction.

It is of interest to compare the egg-box structures of Ca^{2+} and Na^+ as depicted in Fig. 2a and b, respectively. The Ca^{2+} ion may be seen to sit noticeably higher in the binding pocket than the Na^+. This is reflected in the selected relative distances and coordination angles provided in Table 1 and, at first glance, might be considered to be counterintuitive – given that both ions have very similar ionic radii and Ca^{2+} has double the charge of Na^+. However, it should be noted that, relative to Na^+, the binding of the Ca^{2+} is dominated by an interaction with the negatively charged carboxylate donor atom, O'6, (due to its +2 charge). This is reflected in the relative radial distribution functions for the bound Ca^{2+} and Na^+ ions, Fig. 4a and b, respectively. It is clear from these analyses that the Ca^{2+} is approximately six times more likely to be bound to the O'6 that to the other atoms within the coordination sphere. This is not the case for Na^+, where there is a more symmetric binding arrangement within the coordination sphere, Fig. 2. This has the effect of raising the Ca^{2+} ion within the pocket since the carboxylate moiety is orientated vertically. This effect is also enhanced by the presence of a water bridge from the Ca^{2+} to the O'5 atom rather than a direct interaction. This seemingly subtle phenomenon suggests an explanation, at the molecular level, as to why Na^+, unlike Ca^{2+}, does not take part in metal mediated chain aggregation, since, unlike Ca^{2+}, it sits too low into the binding pocket to be accessible to another strand. This is coupled with the fact that Na^+, due to its more even affinity with all the oxygens in the coordination environment, including the carboxylate, Fig. 2, is more likely to associate with poly-G *via* such binding pockets than to form inter-strand carboxylate bridges (as is observed for Ca^{2+} in this study).

4.2. 2 x Poly-G

When this system is modelled with Na^+ ions in the surrounding environment, several more transient instances of the same egg-box binding modality identified previously, Fig. 2, are observed in both of the individual chains. A number of fleeting interactions with single carboxylate groups are also observed. At no point during the simulation did the two chains show signs of Na^+-mediated aggregation, which is consistent with experimental observations regarding the solubility of alginate in the presence of low concentrations of monovalent cations (Donati & Paoletti, 2009).

When a simulation involving two poly-G chains in the presence of Ca^{2+} is carried out, an interesting phenomenon is observed. The two poly-G chains are seen to aggregate almost perpendicular to each other, Fig. 5, with the junction zone being mediated by no less than three calcium ions.

This novel arrangement incorporates an egg-box type Ca^{2+} binding modality (highlighted), which is very similar to the interaction observed with Ca^{2+} and the single poly-G chain in Section 4.1, Fig. 2a. This 'pocket' interaction is reinforced by two interchain carboxylate-Ca^{2+}-carboxylate bridges adjacent to the egg-box interaction and by an interchain hydrogen bond, Fig. 5.

More specifically, a Ca^{2+} ion is embedded into a pocket between two G units and coordinates to four oxygen atoms (O1, O2, O3 and O'6) on one of the chains. The second poly-G chain then interacts with this embedded Ca^{2+} ion *via* a carboxylate oxygen, O6*, with three water molecules completing the coordination sphere. The chain aggregation is further enhanced by an interchain hydrogen bond between the hydroxyl hydrogen on the coordinated O3 oxygen (of the first chain) and the O3 atom one residue removed from the binding site of the second chain, and by the aforementioned two Ca^{2+} ions that form concomitant interchain bridges between carboxyl groups, Fig. 5. This binding junction, involving three Ca^{2+} ions in two different binding modes and defining two poly-G sequences in an approximate perpendicular arrangement, is new to the literature. In terms of the rigidity (stability) of the motif, it should be pointed out that the ratio of residues to calcium ions is 2:1, the same as in the parallel "egg-box" junction model (Grant et al., 1973). Three Ca^{2+} ions at junctions of this type would be entirely consistent with the observed stability of such gels. This motif appears to be quite stable, forming and subsisting for the final 0.35 ns of simulation time, albeit with a slight fluctuation in the intermolecular hydrogen bond, Fig. 6.

The distinctive perpendicular motif described above, is a significant finding since it appears to be ubiquitous in AFM images of open 3-D alginate networks (Decho, 1999). Furthermore, due to the fact that this orientation involves non-terminal G residues, it might be expected that this could occur regardless of the length of the alginate chain. This observation does not imply that G-rich regions of alginate cannot form parallel associations or that they always aggregate perpendicularly; however, these results do demonstrate that, under certain conditions, cross-linking type interactions such as this *may* occur between G regions of alginates. It is worth emphasising that this aggregation is mediated by no less than three Ca^{2+} ions, one of which utilises a geometry similar to the shifted egg-box model (Braccini & Pérez, 2001). These results support the notion that it is possible for G-rich regions to form not only strong parallel associations but also strong *perpendicular* interchain associations. Furthermore, the two supporting interchain carboxylate-Ca^{2+} bridges concomitant to the embedded Ca^{2+}, plus the interchain hydrogen bond would be expected to provide a degree of rigidity to this perpendicular arrangement by decreasing the range of possible movements of the poly-G chains.

4.3. 3 x Poly-G

When a third poly-G decamer is added, in the presence of neutralising Ca^{2+} ions, the resultant aggregation is consistent with previously published theoretical models (Braccini & Pérez, 2001; Grant et al., 1973; Plazinski, 2011). Here, the three chains self-organize approximately parallel, with two of the chains associating more strongly than the third. Thus the two closer chains share five metal-mediated interchain interactions of the

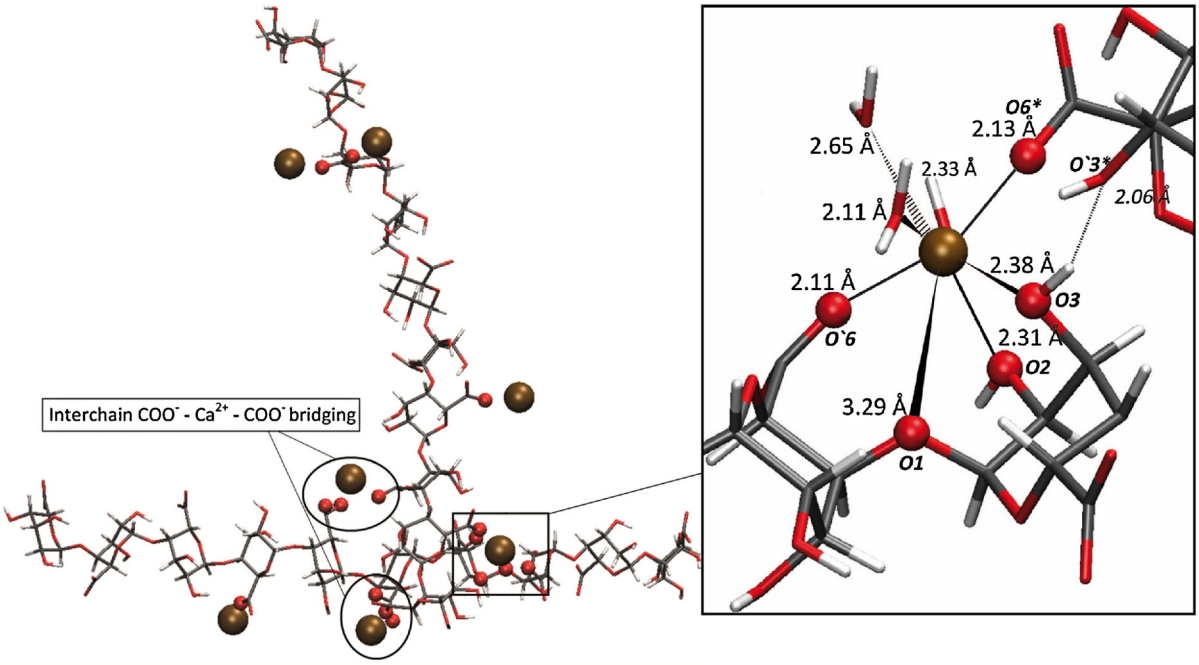

Fig. 5. Two poly-G chains aggregating in a perpendicular motif, *via* three bridging Ca^{2+} ions (brown spheres). Inset highlights the details of the egg-box component of this junction zone. The two buttressing Ca^{2+} interchain carboxylate bridges are also shown, as is the interchain hydrogen bond. Oxygen atoms that are directly interacting with a calcium ion are rendered as small red spheres. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

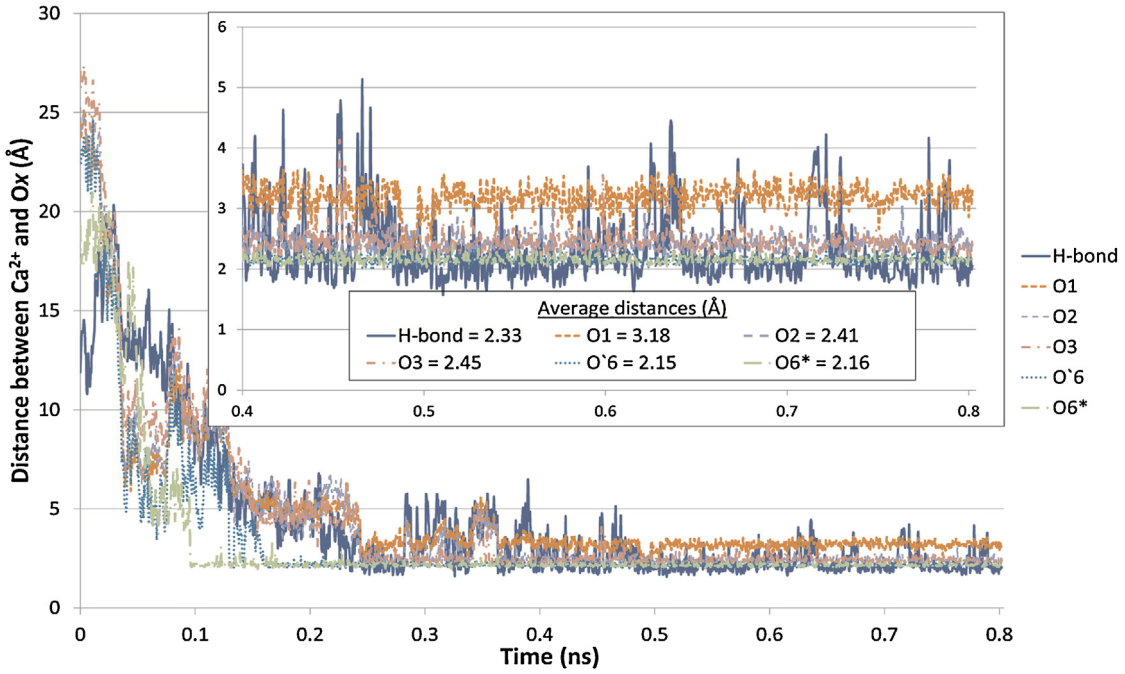

Fig. 6. A plot of the oxygen-Ca^{2+} distances of the atoms involved in the egg-box binding site depicted in Fig. 5. Also shown is the interchain hydrogen bond that is formed between a coordinating water molecule and the O‘3* atom.

carboxylate bridging type – although two sites also invoke binding from a nearby O3 atom, Fig. 7. The third chain associates at only one metal-mediated interaction site, which is *via* a carboxylate bridge. As further evidence that the Ca^{2+} mediated binding of the poly-G chains occurs *via* a variety of mechanisms, rather than by the egg-box model exclusively, this simulation shows the Ca^{2+} ions to interact almost exclusively *via* consolidated carboxylate bridging interactions, similar to the model previously proposed by Plazinsky in 2011. Whilst one egg-box site is identified, this site is disrupted due to steric influences from the second chain attempting to interact through a carboxylate group. Although not as conducive to the formation of open 3-D networks as the previously described perpendicular motif, a parallel aggregation of strands (thickening) with associated branching is also suggested by available AFM images of such networks (Decho, 1999).

To what extent these structures represent intermediate or conserved motifs in the assembly mechanism would obviously require more enquiry. However, it may be surmised that, with

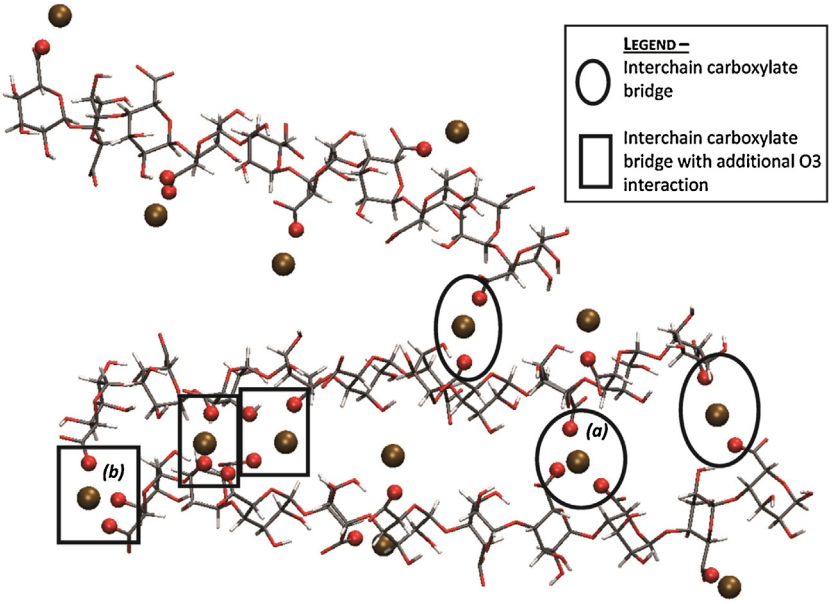

Fig. 7. Three poly-G chains aggregating in an approximately parallel arrangement in the presence of Ca^{2+} ions (shown in brown). Oxygen atoms directly interacting with the Ca^{2+} ions are shown as red spheres. The interchain interaction labelled (a) is a carboxylate bridge with three carboxylate oxygens contributing, and (b) involves an O4 atom rather than an O3; however, this is due to an end-chain effect. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

further simulation time, the 3 x poly-G parallel arrangement, in particular, could relax into an egg-box coordination pattern. Such relaxations, from metal-mediated carboxylate bridging to consolidated egg-box-type binding sites, might suggest a molecular basis for the experimentally observed syneresis identified in alginate hydrogels (Donati et al., 2005; Donati & Paoletti, 2009).

5. Conclusions

As described above, the MD method and simulations presented here have resulted in the characterisation of a variety of intrinsic binding modes for the interaction of sodium and calcium ions with single and multiple poly-G decamers that have implications for chain aggregation. All the coordination geometries, in terms of coordination numbers, bond lengths and angles, are well-behaved and as expected for the ions studied, which is a strong validation for the incorporation of the simulated annealing protocol. A slight variant on the classical egg-box binding mode was revealed with respect to Ca^{2+} binding and an analogous (though not identical) mode for Na^{+} has been characterised for the first time. A detailed comparison of the coordination spheres of these two ions provides a compelling structural rationale for the inability of Na^{+} compared to Ca^{2+} to induce chain aggregation - as is observed both experimentally and *via* simulation (this study). Several potential motifs have been uncovered that are consistent with previously reported molecular-scale imaging of an alginate gel network formed in the presence of such ions. Particularly noteworthy is the discovery of a stable perpendicular motif, mediated by three Ca^{2+} ions, that appears to be ubiquitous in such images and that could provide an important structural basis for the assembly of open 3-D networks. Overall, direct interactions of both ions with single carboxylate moieties are common and water-bridging interactions from the metal to oxygen atoms are also observed, as is inter-chain hydrogen bonding. This study provides significant insights into the intrinsic interactions of sodium and calcium ions with poly-G and how such interactions might underpin the assembly of open 3-D gels.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.carbpol.2013.11.034.

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